Haematological effects of aqueous extract of Ornamental plants in male Swiss albino mice

N Mishra* and V L Tandon

Department of Bioscience and Biotechnology, Banasthali University Banasthali,Rajasthan-304022, India * Corresponding author email: nidhi.mishraa12@gmail.com Received:26-07-2011, Accepted:09-08-2011, Published Online: 17-11-2011 doi: 10.5455/vetworld.2012.19-23

Abstract

Treatment of mice with crude extract of *Hibiscus rosa sinensis* flowers (500 mg/kg BW) and *Bougainvillea spectabilis* leaves (800 mg/kg BW) for a period of 30 days indicates a significant increase in the level of hemoglobin and count of RBC but a significant decline in the level of MCH and MCV in the former case. On the other hand, in *B. spectabilis* treated animals, the level of hemoglobin, RBC count & PCV declined significantly. Hence, it is concluded that the use of *H. rosa sinensis* whereas may not cause any adverse effect on animals, *B. spectabilis* is to be used with care as its chronic use may cause anemia.

Keywords: Crude extract, H. rosa sinensis, B. spectabilis, hemoglobin, MCH, MCV, PCV

Abbreviations: BW (Body Weight), Hb (Haemoglobin), MCV (Mean Corpuscular Volume), MCH (Mean Corpuscular haemoglobin), MCHC (Mean Corpuscular Haemoglobin Content), PCV (Packed Cell Volume), PLT(Platelets), RBC (Red Blood Cells), WBC (White Blood Cells)

Introduction

For thousands of years, people have looked to natural means of healing. In developing countries of the world, most the people depend on herbal medical care (Ekpe et al., 1990). The popularity of traditional medicine is due to the belief that some diseases only respond to traditional treatment (Bannerman et al., 1975). Extracts of root, stem, bark, leaves and flowers of some medicinal plants have been shown to have activities against most dreaded pathogenic organisms like the bacteria, fungi etc (Bannerman et al., 1975; Khan et al., 1980; Madunagu et al., 1990; Singh and pattak, 1994), while some others are cytotoxic (Russel et al., 1997; Prohp and Alaiya, 2003; Prohp and Maduduemezia, 2004; Prohp et al., 2004, Prohp et al., 2006a, Prohp et al., 2006b). Besides, ingestion of some plant materials (either in the raw form or their extracts) having useful medicinal properties may cause anaemia resulting from the sequestration of RBC in spleen, impaired red blood cell production or primary bone marrow dysfunction (Watt and

Brever, 1962; Cheeke, 1998).

In the present study, an attempt is made to study the therapeutic value of the crude extract of two most common locally available ornamental plants flourishing throughout the year, namely *Hibiscus rosa sinensis* and *Bougainvillea spectabilis*. Keeping in mind the report of Williamsons (2001), who has emphasized that the whole or partially purified extract of plant offer advantages over a single isolated ingredient.

In this investigation, we have sought to verify the effect of the administration of aqueous extract of *H. rosa sinensis* flowers and *B. spectabilis* leaves on few hematological parameters.

Materials and Methods

Animals: Adult male and female Swiss albino mice procured from CCS Haryana Agricultural University, Hisar were mated and resulting progeny was maintained in a well ventilated animal house with 12:12 light/dark cycle. After quarantine and acclimization period of 1 week mice were used for this study.

Test animals were kept in polyproline cages

with iron bar tops and maintained on standard pellet diet (Hindustan Levers). Tap water was made available *ad libitum*. As far as possible, necessary sterile conditions were provided and cleanliness was maintained in the animal cages as well as in the room. Prior approval for experiments was taken from Institutional Animal Ethics Committee as per CPCSEA (Govt. of India) norms.

Preparation of dose formulations: Extraction of aqueous extract of *H. rosa sinensis* and *B. spectabilis* the Flowers and leaves collected from Banasthali University campus were oven dried and powdered. The powdered material was extracted using aqueous by soxhlet method (Sharma and Grag, 2009). It was further dried, which was used for the experimentation.

Experimental Protocol: The animals were randomly selected into 3 groups and each group contained 12 animals each:

Ist Group: Treated orally with aqueous *H. rosa* sinensis flowers extract per day at the dose of 500mg/kg BW for 30 days.

Hnd Group: Treated orally with aqueous *B. spectabilis* leaves extract per day at the dose of 800mg/kg BW 30 days.

IIIrd Group: Control group.

Blood was withdrawn from eye orbit at regular interval of 10th, 20th, 30^{th} day and after withdrawal of the treatment for 60 days.

Collection of samples: For haematological parameter blood sample was collected from retro – orbital plexus using micro – capillary technique (Sorg and Buckner, 1964). Heparin was used as an anticoagulant.

Determination of Haematological Parameters: Blood was collected into EDTA-containing sample bottles on day 10th, 20th, 30th and 90th from experimental animals for hematological analysis as recommended by Malomo *et al.*, (2002). RBC and WBC counting was done with the help of Neubaur's chamber. Packed cell Volume (PCV) with Wintrobe hematocrit tubes, Haemoglobin (Hb) by Sahli's method (Sharma, 2007). Other haematological indices were calculated with the aid of formulas. MCV (Mean Corpuscular Volume) - The average volume of RBC. Expressed in femtoliters (cubic micrometers).

MCH (Mean Corpuscular haemoglobin) -The average content (mass) of Hb in a given volume of packed RBCs, expressed in picograms.

MCHC (Mean Corpuscular Haemoglobin Content) is the average concentration of Hb in a given volume of packed RBCs, expressed in grams per deciliter or in percent.

Statistical Analysis: Results are expressed as mean \pm Standard Deviation (S.D.). Statistical significance between the different groups was determined by one way Analysis of Variance (ANOVA) using the SPSS (Ver. 16). Post hoc testing was performed for inter-group comparisons using the Tukey multiple comparison test at P<0.05. Whenever sphericity was significant, degree of freedom and F-value are corrected by Huynh Feldt epsilon.

Results

Effect on Haematological parameters: The effect of aqueous extract of *H. rosa sinensis* flowers and *B. spectabilis* leaves on Swiss albino mice on day 10^{th} , 20^{th} and 30^{th} along with a recovery period are presented in Table 1. Administration of *H. rosa sinensis* aqueous extract produced significant increase (P<0.05) in the count of the RBC other along with Hb content and decline in Platelet count however, no effect could be observed on MCHC level. On the contrary in case of *B.spectabilis* treated animals, the count of RBC declined significantly along with Hb level whereas increase in WBC. However the level of PCV, MCH, MCHC and platelet count remained unaffected.

Discussion

The administration of any chemical compound for long duration may bring about significant changes in the structure, function, metabolic transformation and concentration of biomedical enzymes and even metabolic pathways. These alterations may be rapid or slow and may lead to different biochemical mechanisms, producing a pathological state (Murray *et al.*, 2000). Haematological effects of aqueous extract of Ornamental plants in male Swiss albino mice

(a) Hasmatalagical Parameters
haematological parameters and (b) haematological Indices in male mice.
Table-1. Effect of aqueous extract of <i>H. rosa sinensis</i> flower and <i>B. spectabilis</i> leaves on some (a)

Control Lrosa sinensis B.spectabilis	20.13±0.110 19.97±0.309	20.3±0.374	20.28±0.291	20 15+0 135
Control I.rosa sinensis B.spectabilis Control	20.13±0.110 19.97±0.309 21.6+1.265*	20.3±0.374	20.28±0.291	20 15+0 135
Lrosa sinensis S.spectabilis Control	19.97±0.309	40 0.0 070		20.1010.100
8.spectabilis Control	01 G 1 0 CE*	19.8±0.676	19.63±0.727	20.47±0.782
Control	21.0±1.303	22.77±1.126*	23.27±0.228*	22.6±1.537*
	9.38±0.31	9.43±0.27	9.33±0.34	9.41±0.28
l.rosa sinensis	9.96±0.47*	10.01±0.14	10.18±0.39*	10.09±0.38*
3.spectabilis	9.28±0.36	9.23±0.41	9.26±0.34	9.36±0.31
Control	13.42±0.375	13.18±0.563	13.35±0.369	13.28±0.480
l.rosa sinensis	13.53±0.984	13.87±0.634	14.2±0.216	13.83±0.541
3.spectabilis	12.32±0.315*	12.33±0.589	10.72±0.978*	12.16±0.637*
ontrol	47.20±1.330	47.03±0.970	47.04±1.497	46.87±1.730
l.rosa sinensis	46.77±0.552	47.93±1.659	48.27±3.369	46.43±1.019
3.spectabilis	42.24±1.394*	40.49±3.561*	39.66±2.761*	42.32±1.312*
ontrol	1253.17±41.862	1254.83±40.321	1254.8±39.486	1254.3±44.187
l.rosa sinensis	905.67±41.624*	879±17.426*	855.66±10.609*	870±22.211*
8.spectabilis	1251.5±43.668	1249.67±45.923	1236.5±35.780	1246.5±44.761
dices				
reatment	10 days	20days	30days	90days
ontrol	14.32±0.45	13.99±0.62	14.33±0.46	14.13±0.56
l.rosa sinensis	13.32±0.48	13.48±1.05	11.56±0.86*	13.01±0.42*
3.spectabilis	13.62±1.33	13.89±0.92	14.05±0.33	13.84±0.92
ontrol	28.44±0.89	28.01±1.07	28.41±1.02	28.37±1.09
l.rosa sinensis	29.22±1.55	30.66±2.84	27.14±2.89	28.76±1.68
3.spectabilis	28.77+2.07	28.95±1.40	29.62+2.00	29.82±1.55
Control	50.36±1.96	49.91±1.63	50.48+2.47	49.82+2.08
l.rosa sinensis	45.61±2.72*	43.86±3.42*	42.85±2.80*	45.31±1.93*
S.spectabilis	47.05±1.83	47.95±1.96	47.46±3.22	46.12+2.28*
	I. rosa sinensis Sepectabilis Sontrol I. rosa sinensis Sepectabilis Sontrol I. rosa sinensis Sepectabilis Sontrol I. rosa sinensis Sepectabilis Sontrol I. rosa sinensis Sepectabilis Sontrol I. rosa sinensis Sepectabilis Sontrol I. rosa sinensis Sepectabilis Sepectabilis Sepectabilis Sepectabilis Sepectabilis Sepectabilis	I. rosa sinensis $9.96\pm 0.47^*$ Aspectabilis 9.28 ± 0.36 control 13.42 ± 0.375 I. rosa sinensis 13.53 ± 0.984 spectabilis $12.32\pm 0.315^*$ control 47.20 ± 1.330 I. rosa sinensis 46.77 ± 0.552 Aspectabilis $42.24\pm 1.394^*$ control 1253.17 ± 41.862 I. rosa sinensis $905.67\pm 41.624^*$ Spectabilis 1251.5 ± 43.668 dices 10 days reatment 10 days control 14.32 ± 0.45 I. rosa sinensis 13.62 ± 1.33 ontrol 28.44 ± 0.89 I. rosa sinensis 29.22 ± 1.55 spectabilis 28.7 ± 2.07 ontrol 50.36 ± 1.96 I. rosa sinensis $45.61\pm 2.72^*$ spectabilis $45.61\pm 2.72^*$	I.rosa sinensis $9.96\pm0.47^*$ 10.01 ± 0.14 I.spectabilis 9.28 ± 0.36 9.23 ± 0.41 I.rosa sinensis 13.42 ± 0.375 13.18 ± 0.563 I.rosa sinensis 13.53 ± 0.984 13.87 ± 0.634 I.spectabilis $12.32\pm0.315^*$ 12.33 ± 0.589 I.rosa sinensis 47.20 ± 1.330 47.03 ± 0.970 I.rosa sinensis 46.77 ± 0.552 47.93 ± 1.659 I.rosa sinensis 46.77 ± 0.552 47.93 ± 1.659 I.rosa sinensis $42.24\pm1.394^*$ $40.49\pm3.561^*$ I.rosa sinensis $905.67\pm41.624^*$ $879\pm17.426^*$ I.spectabilis 1253.17 ± 41.862 1254.83 ± 40.321 I.rosa sinensis $905.67\pm41.624^*$ $879\pm17.426^*$ I.spectabilis 1251.5 ± 43.668 1249.67 ± 45.923 dicesInterment10 days20dayscontrol 14.32 ± 0.45 13.99 ± 0.62 I.rosa sinensis 13.62 ± 1.33 13.89 ± 0.92 ontrol 28.44 ± 0.89 28.01 ± 1.07 I.rosa sinensis 29.22 ± 1.55 30.66 ± 2.84 I.spectabilis 28.77 ± 2.07 28.95 ± 1.40 ontrol 50.36 ± 1.96 49.91 ± 1.63 I.rosa sinensis $45.61\pm2.72^*$ $43.86\pm3.42^*$ I.rosa sinensis $45.61\pm2.72^*$ $43.86\pm3.42^*$	I.rosa sinensis $9.96\pm0.47^*$ 10.01 ± 0.14 $10.18\pm0.39^*$ 1.spectabilis 9.28 ± 0.36 9.23 ± 0.41 9.26 ± 0.34 1.ontrol 13.42 ± 0.375 13.18 ± 0.563 13.35 ± 0.369 I.rosa sinensis 13.53 ± 0.984 13.87 ± 0.634 14.2 ± 0.216 1.spectabilis $12.32\pm0.315^*$ 12.33 ± 0.589 $10.72\pm0.978^*$ 1.rosa sinensis 46.77 ± 0.552 47.93 ± 1.659 48.27 ± 3.369 1.rosa sinensis 46.77 ± 0.552 47.93 ± 1.659 48.27 ± 3.369 1.spectabilis $42.24\pm1.394^*$ $40.49\pm3.561^*$ $39.66\pm2.761^*$ 1.ontrol 1253.17 ± 41.862 1254.83 ± 40.321 1254.8 ± 39.486 1.rosa sinensis $905.67\pm41.624^*$ $879\pm17.426^*$ $855.66\pm10.609^*$ 1.spectabilis 1251.5 ± 43.668 1249.67 ± 45.923 1236.5 ± 35.780 dicesreatment 10 days 20 days30 dayscontrol 14.32 ± 0.45 1.rosa sinensis 13.62 ± 1.33 13.89 ± 0.92 14.05 ± 0.33 ontrol 14.32 ± 0.45 13.99\pm0.62 14.33 ± 0.46 I.rosa sinensis13.32\pm0.48 13.48 ± 1.05 $11.56\pm0.86^*$ spectabilis13.62\pm1.33 13.89 ± 0.92 14.05 ± 0.33 ontrol 28.44 ± 0.89 28.01 ± 1.07 28.41 ± 1.02 I.rosa sinensis 29.22 ± 1.55 30.66 ± 2.84 27.14 ± 2.89 spectabilis 28.77 ± 2.07 28.95 ± 1.40 29.62 ± 2

The values are expressed as mean \pm SD for 10 animals (n=10) per group.

Treatment was discontinued after 30 days, and animals were sacrificed 60 days after withdrawal of treatment.

^{*} indicates statistically significant with Control at p < 0.05.

In the present study, aqueous extract of *H.* rosa sinensis and *B.* spectabilis has investigated on hematological parameters on day 10, 20 and 30 of treatment. The results obtained are summarized in table 1a & b. The results indicate, that the treatment with *H.* rosa sinensis for 30 days significantly increases the count of RBC and WBC from day 10 onward but causes a significant decline in the count of blood platelets. Besides, through Hb, PCV, MCHC remain unaffected, MCH decreased significantly.

However, in the case of *B. spectabilis* extract treated animals for the same duration, though WBC count, MCH, MCHC, MCV and PLT remain unaffected, a significant decline in RBC and PCV is indicated.

Actually, assessments of haematological parameters are used to determine the extent of deleterious effect of the extracts on blood of an animal. Straus, (1998), Adedapo *et al.*, (2007), Onyeyilli *et al.*, (1998) reported, that reduction in RBC, Hb and PCV is an indication of either the

destruction of RBC or their decreased production, which may lead to anemia.

On the contrary an increase in the count of RBC, Hb and PCV is suggestive of polycythemia and positive erythopoisis (Iranloye, 2002; Mansi and Lahham, 2008; Kuppast *et al.*, 2009; Okpuzor *et al.*, 2009). Hence, a significant increase in RBC with no alteration in Hb and PCV in *H. rosa sinensisi* treated animals indicates that the extract causes no toxic effect on RBC. However, a significant reduction of RBC and Hb from day 10 in *B. spectabilis* treated animals is suggestive of anaemia.

Reports about WBC counts have pointed out that whereas increased count of WBC is supposed to be helpful in boosting immune system (Adedapo *et al.*, 2007; Mohajeri, 2007), a decreased count of WBC shows the suppression of leucocytes and their production from bone marrow (Odesanmi *et al.*, 2010; Jimoh *et al.*, 2008; Osuigwe *et al.*, 2007; Adedapo *et al.*, 2004).

Therefore, an increased count of WBC in *B*.

spectabilis treated animals, as observed in the present study, suggests that *B. spectabilis* might be having a good potentiality to boost immune system. However, this report in not agreement with Adebayo *et al.*, (2005), where they have reported that ethanolic extract of *B. spectabilis*, even at the dose of 200mg/kg, decreases the count of WBC.

Blatti, (1997), Adedapo *et al.*, (2008), Adeniyi *et al.*, (2010)have pointed out, that reduced blood platelets affect the viscosity of blood, which is corelated positively to blood pressure. As shown in the table, *H. rosa sinensis* treatment for 30 days adversely affects the count of blood platelets which may produce negative effect on the viscosity of blood. Probably the duration of the treatment may prove toxic to animals.

The hematological parameters like MCH, MCHC and MCV are related to individual RBC, while Hb and PCV are associated with total population of RBCs. Therefore, if MCH, MCHC and MCV are not affected by the treatment with extracts, it means that neither the incorporation of Hb into RBC, nor the morphology and osmotic fragile of RBCs is altered (Adebayo *et al.*, 2005; Ashafa *et al.*, 2009).

It is thereby concluded, that both the extracts are selectively toxic to hematological parameters. Chronic use of the extracts is not advisable, despite of their popularity having antidiabetic and antilipidemic properties.

Acknowledgment

Authors are thankful to Professor Aditya Shastri for kindly extending "Banasthali Centre for Education and Research in Basic Science" sanctioned under CURIE (Consolidation of University Research for Innovation and Excellence in Women University) programme of department gratefully acknowledged. Nidhi Mishra also expressed her gratitude to Department of Science and Technology, New Delhi for financial assistance (WOS-A Project No. SR/WOS-A/LS-41/2009).

Conflict of interest

Authors declare that they have no conflict of interest.

www.veterinaryworld.org

References

- 1. Adebayo, J. O., Adesokan, A. A., Olatunji, L. A., Buoro, D. O. and Soladoye, A. O, (2005). Effect of ethanolic extract of Bougainvillea spectabilis leaves on haematological and serum lipid variables in rats. *Biokemistri*. 17: 45-50.
- 2. Adedapo, A. A., Abatan, M. O. and Olorunsogo, O. O, (2004). Toxic effects of some plants in the genus Euphorbia on haematological and biochemical parameters in rats. *Veterinarski Arhiv*. 74: 53-62.
- Adedapo, A. A., Abatan, M. O, and Olorunsogo, O. O, (2007). Effect of some plants of the spurge family on haematological and biochemical parameters in rats. *Veterinarski Arhiv.* 77: 29-38.
- 4. Adedapo, A. A., Sofidiya, M. O., Masika, P. J. and Afolayan, A. J. (2008). Safety evaluations of the aqueous extract of Acacia karroo stem bark in rats and mice. *Records of Natural Products*. 2; 128-134.
- 5. Adeniyi, T. T., Ajayi, G. O., Akinsanya, M. A. and Jaiyeola, T. M, (2010). Biochemical changes induced in rats by aqueous and ethanolic corm extract of *Zygotritonia croceae*. *Scientific Research and Essay.* 5; 71-76.
- 6. Ashafa, A. O. T., Yakubu, M. T, Grierson, D. S. and Afolayan, A. J, (2009). Effects of aqueous extract from the leaves of *Chrysocoma ciliate L*. on some biochemical parameters of wistar rats. *Afr. J. Biotech.* 8: 1425-1430.
- Bannerman, R. H. A., Ummina, V. D. and Koko, U, (1975). Indigenous system of medicine in India. In: Alternative Approaches to Meeting basic Health needs in developing countries, *WHO*, *Geneva*, 84-19.
- 8. Cheeke, P.R, (1998). Natural toxicants in feeds, forages and poisonous plants.2nd Edition. IL: Interstate Publishers, Danville.
- Ekpe, E. D., Ebana, R. V. B. and Madunagu, B.E, (1990). Antimicrobial activity of four medicinal plants on pathogenic Bacteria and phytopathogenic fungi. *West Afr. J. Biological Appl Chem.* 35: 2-5.
- 10. Iranloye, B. O, (2002). Effect of chronic garlic feeding on some haematological parameters. *Afr. J. Biomedical Res.* 5: 81-82.
- Jimoh, O. R., Olaore, J., Olayaki, L. A., Olawepo, A. and Biliaminu, S. A, (2008). Effects of aqueous extract of *Ocimmum gratissimum* on haematological parameters of wistar rats. *Biokemistri*. 20; 33-37.

- Khan, M. R., Nddaalio, G., Nkunja, M. H., Weever, H. and Sawhney, A. H, (1980). Studies on the African Medicinal plants part 1. Preliminary screening of medicinal plant for antifungal activity. *Plant Med. Suppl.* 40: 91-92.
- 13. Kuppast, I. J., Vasudeva, Nayak, P., Ravi, M. C. and Biradar, S. S. (2009). Studies on the hematological effect of the extracts of Cordiadichotoma Forst. F. Fruits. *Res. J. Pharmacol. Pharamacodynamics*. 1: 117-119.
- Madunagu, B. E., Ebana, R. U. B. and Ekpe, E. D, (1990). Antibacterial and Antifungal Activity of some medicinal plants of Akwa Ibom state. *West Afr. J. Biolological Appl. Chem.* 35: 25-30.
- 15. Malomo, S. O., Adebayo, J. O., Olorunniji, F. J, (2002). Modulatory effect of vitamin E on some haematological parameters in dihydroartemisinin-treated rats. *Trop. J. Health Sci.* 9: 15-20.
- 16. Mansi, K. and Lahham, J, (2008). Effects of Artemisia sieberi Besser (A/ herba-alba) on heart rate and some hematological values in normal and alloxan induced diabetic rats, *J. Basic and Appl. Sci.* 4: 57-62.
- 17. Mohajeri, D., Mousavi, G. and Mesgari, M, (2007). Subacute toxicity of Crocus sativus L. (Saffron) stigma ethanolic extracts in rats. *Am. J. Pharmacol. Toxicol.* 2: 189-193.
- Murray, R. K., Granner, P. A., Mayer, P. A. and Rodwell, V. W, (2000). Harper's Biochemistry. 20th edition. McGraw-Hill, 594-602.
- 19. Sharma, N and Garg V, (2009). Antihyperglycemic and antioxidative potential of hydroalcoholic extract of Butea monosperma Lam flowers in alloxan-induced diabetic mice. *Ind. J. Exp. Biol.* 47: 571-574.
- Odesanmi, S. O., Lawal, R. A. and Ojokuku, S. A, (2010). Haematological effects of ethanolic fruit extract of Tetrapleura tetraptera in male Dutch white rabbits. *Res J. Med. Plant.* 4: 213-217.
- 21. Okpuzor, J., Ogbunugafor, H. A. and Kareem, G. K. (2009). Hepatoprotective and hematologic effects of fractions of Globimetula braunii in normal albino rats. *EXCIL Journal*. 8: 182-189.
- 22. Osuigwe, D. I., Nwosu, C. and Oguni, J. O, (2007). Preliminary observations on some haematological parameters of juvenile heterobranchus longifilis fed different dietary level of Raw and boiled jackbean (Canavalea ensiformis) seed meal. Conference on International Agricultural Research for Development, 1-6.
- 23. Prohp, T. P and Alaiya, H. T, (2003). Some functional properties and anti-nutritional factors

www.veterinaryworld.org

Veterinary World, Vol.5 No.1 January 2012

of extracotyledonous deposits of pride of Barbados (*Caesalpina pulcherrima*). Proceedings (15th Annual conference of BSN held in AAU, Ekpoma, 2002), 40-45.

- 24. Prohp, T. P and Maduemezia, C, (2004). Carbohydrate, ash and moisture contents of extra-cotyledonous deposits of pride of Barbados (*Caesalpina pulcherrima*). *Niger. J. Agric. Sci. Forestry.* 1: 195-2004.
- 25. Prohp, T. P., Ihimire, I. G., Madusha, A. O., Okpala, H. O., Erebor, J. O. and Oyinbo, C. A, (2006a). Some antinutritional and mineral contents of extracotyledonous deposits of pride of Barbados (*C.pulcherrima*). *Pak. J. Nutr.* 5: 114-116.
- Prohp, T. P., Mendie, E. A., Madusha, A. O., Uzoaru, S. C., Aigbiremolen, A. and Onyebuagu, P. C, (2004). Cyanide contents of pride of Barbados (*Caesalpina pulcherrima*) grown in different parts of *Niger. J. Med. Laboratory Sci.* 13: 29-32.
- Prohp, T. P., Osifo, E. S, Madusha, A. O., Erebor, J. O., Okpala, H. O. and Oyinbo, C. A, (2006b). Effects of aqueous extract of extra-cotyledonous deposits of pride of Barbados (*Caesalpina pulcherrima*) on some blood electrolytes and urea levels in rabbits. *Pak. J. Nutr.* 5: 239-241.
- Russel, B. A., Hardin, J. N., Grand, L. and Traser, A, (1997). Poisonous plants of North Carolina, caesalpina sp (pride of Barbados). Department of Horticultural science. North Carolina state University (on line) http://www. ces.nesy.edu/depths.
- 29. Sharma, S, (2007). Experiments and techniques in Biochemistry, Haematology determination. Galgotia, (New Delhi), 90-100.
- Singh, K. V. and Pattak, (1994). Effects of leaf extract of some higher plants on spore germination of Usilago Maydis and U. nuda. Fitoterapia. 55: 318-320.
- 31. Sorg, D. A and Buckner, B, (1964). A simple method of obtaining venous blood from small laboratory animals. *Proceedings of the Society for Experimental Biology and Medicine*, 115: 1131-1132.
- 32. Straus, J.H, (1998). Anaemia. In: Merck Veterinary Manual: A handbook of diagnosis, and therapy for Veterinarians. 8th ed. Merck and Co. Inc. Whitehouse Station, N. J. U.S.A, 8-18.
- Watt, J. M. and Breyer, B, (1962). The medicinal and poisonous plants of Southern and East Africa. 2nd Edition. E. & S. Livingstone Ltd., Edinburgh.

23